

## Forum Review

# NADPH Oxidase-Mediated Oxidative Stress: Genetic Studies of the *p22<sup>phox</sup>* Gene in Hypertension

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### ABSTRACT

Increased vascular production of reactive oxygen species, especially superoxide anion, significantly contributes to the oxidative stress associated with hypertension. An enhanced superoxide production causes an increased inactivation of nitric oxide that diminishes nitric oxide bioavailability, thus contributing to endothelial dysfunction and hypertrophy of vascular cells. It has been shown that NADPH oxidases play a major role as the most important sources of superoxide anion in phagocytic and vascular cells. Several experimental observations have described an enhanced superoxide generation as a result of NADPH oxidase activation in hypertension. Although these enzymes respond to stimuli such as vasoactive factors, growth factors, and cytokines, recent data suggest a significant role of the genetic background in the modulation of the expression of its different components. Several polymorphisms have been identified in the promoter and in the coding region of *CYBA*, the gene that encodes the essential subunit of the NADPH oxidase *p22<sup>phox</sup>*, some of which seem to influence significantly the activity of these enzymes in the context of cardiovascular diseases. Among *CYBA* polymorphisms, genetic investigations have provided a novel marker, the  $-930^{A/G}$  polymorphism, which determines the genetic susceptibility of hypertensive patients to oxidative stress. *Antioxid. Redox Signal.* 7, 1327–1336.

### OXIDATIVE STRESS

THE TERM “OXIDATIVE STRESS” refers to the situation in which cells are exposed to high concentrations of molecular oxygen or its derivatives, also known as reactive oxygen species (ROS). ROS include free radicals, such as superoxide anion ( $\cdot\text{O}_2^-$ ), hydroxyl radical ( $\cdot\text{OH}$ ), nitric oxide ( $\cdot\text{NO}$ ), and lipidic radicals, and other remarkably reactive molecules, such as hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) and peroxynitrite ( $\text{ONOO}^-$ ) (20, 45). Due to their aggressive oxidative action, ROS were considered to be mere toxic agents, as they oxidize macromolecules, including DNA, proteins, and lipids. In recent years, the pathophysiological role of ROS as second messengers has been highlighted in the literature.

The balance between redox homeostasis and oxidative stress is delicate. In physiological conditions, the changes in

type and levels of ROS are finely regulated by the strict control of the enzymatic sources and by proper ROS-scavenging actions of antioxidant systems. Thus, it is due to the excess in ROS production and/or the lack of sufficient defense that ROS are notably increased in pathological situations. This redox imbalance leads to the plethora of disorders in which oxidative stress is known to be involved. ROS take part in aging and senescence, malignant processes, neurodegenerative diseases, rheumatoid arthritis, diabetes, and cardiovascular diseases, including atherosclerosis and hypertension (20).

The implication of oxidative stress is known in a wide number of models of cardiovascular disorders and human cardiovascular disease (29). For instance,  $\text{H}_2\text{O}_2$  levels seem to be increased (49) and superoxide dismutase levels diminished (47) in human hypertension. Oxidative stress also participates in several of the mechanisms involved in atherosclerosis (34,

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48). Besides, clinical trials of antioxidants have had some success in the treatment of cardiovascular diseases (74, 80).

The cardiovascular system is a perfect example of the dual effect of ROS (30). At physiological levels, ROS participate in vascular and cardiac cell function, as they regulate processes ranging from gene control to cell-to-cell interactions, including vascular apoptosis, expression of adhesion molecules, matrix regulation, and migration (38, 81). Two of the key vascular processes regulated by oxidative stress in hypertension and atherosclerosis are the control of vessel tone and the control of vascular cell growth. Therefore, pathological levels of ROS lead to the imbalance of cell function and cause, among others, endothelial dysfunction and vascular hypertrophy.

The depletion of  $\cdot\text{NO}$ , the main vascular relaxing factor, by its reaction with  $\cdot\text{O}_2^-$  leads to an impaired relaxation of the vessel, characteristic of endothelial dysfunction (10, 45). Besides, the  $\text{ONOO}^-$  generated by their reaction is another potent oxidant (6). On the other hand,  $\text{H}_2\text{O}_2$ , product of the dismutation of  $\cdot\text{O}_2^-$ , constitutes a key signal for vascular hypertrophy (38, 55).

The sources of ROS are diverse, present in many cells and tissues. They include the mitochondrial electron transport system, cytochrome P450, lipoxygenase, cyclooxygenase, peroxidases, hemoproteins, xanthine oxidase,  $\cdot\text{NO}$  synthase, and NADPH oxidases (45). Nonetheless, in the last decade, the study of vascular oxidative stress has provided multiple evidences of the critical involvement of the NADPH oxidase system in the molecular basis of cardiovascular diseases, including hypertension (31, 93).

## THE NADPH OXIDASE SYSTEMS

This membrane-associated enzyme system was first described in phagocytic cells, where it plays a role in immune protection due to its bactericidal activity (13). However, it is also present in vascular cells [reviewed by R. Touyz in this issue (82)], where its function is not well known, although it has been hypothesized to play a role in host defense, signal transduction, oxygen sensing, and metabolism (50). These phagocytic and vascular NADPH oxidases are related, but have

structural and functional differences. The NADPH oxidase was well characterized in phagocytic cells, where there are two membrane-bound elements ( $\text{gp91}^{\text{phox}}/\text{NOX-2}$  and  $\text{p22}^{\text{phox}}$ ), three cytosolic subunits ( $\text{p47}^{\text{phox}}$ ,  $\text{p67}^{\text{phox}}$ , and  $\text{p40}^{\text{phox}}$ ), and a low-molecular-weight G protein ( $\text{rac1}$  or  $\text{rac2}$ ) (13). The vascular NADPH oxidase is similar to the phagocytic oxidase, but there is controversy about the expression of several of its components, depending on cell type and species considered (Table 1). Besides, in the last years, several homologues have been discovered for these subunits (82). These structural differences among vascular and phagocytic NADPH oxidases could help explain their functional differences. In this way, phagocytic oxidases have an inducible activity with a large, extracellular  $\cdot\text{O}_2^-$  production, whereas vascular oxidases are constitutive enzymes that produce small amounts of  $\cdot\text{O}_2^-$  (52).

Several events take part in the phagocytic NADPH oxidase activation, such as phosphatidylinositol phosphates accumulation, interactions with cytoskeleton components, and phosphorylation of several subunits (13). The activation of vascular NADPH oxidases is not as well known as that of the phagocytic oxidase, but the mechanism is likely to be similar. In fact,  $\text{p47}^{\text{phox}}$  phosphorylation is key to activation of phagocytic and vascular NADPH oxidases (13, 52). Several signaling pathways activate both the phagocytic and the vascular NADPH oxidases, such as protein kinase C activation, phospholipase D pathway, and phospholipase  $\text{A}_2$  pathway (82).

Although vascular NADPH oxidases are constitutive enzymes, several stimuli, including humoral factors and hemodynamic forces, have been shown to regulate their activity (82). On the other hand, phagocytic NADPH oxidase is activated, among others, by nonopsonized zymosan, phorbol myristate acetate, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interferon- $\gamma$ , and interleukin-15 (22, 66).

## NADPH OXIDASES IN HYPERTENSION

Nowadays, the central role of NADPH oxidases in cardiovascular diseases such as hypertension (53, 93), diabetes (33, 36), atherosclerosis (34), and cerebrovascular disease (65) is widely known. This role has been determined in experimental

TABLE 1. EXPRESSION OF NADPH OXIDASE COMPONENTS

Components	Tissues or cell types	References
Nox1	Colon, VSMCs, ECs, fibroblasts	42, 43, 78, 87
Nox2/ $\text{gp91}^{\text{phox}}$	Neutrophils, macrophages, ECs, VSMCs, fibroblasts, cardiomyocytes	5, 13, 41, 46, 87
Nox 3	Fetal kidney, lung, spleen, and liver	3, 11
Nox 4	Kidney, heart, osteoclasts, pancreas, placenta, VSMCs	3, 11, 27, 42, 77, 78, 90
Nox 5	Testis, spleen, lymph nodes, ovary, placenta, pancreas, human VSMCs	2, 3, 11
Duox1	Thyroid	16, 21
Duox2	Thyroid, colon	16, 21
$\text{p22}^{\text{phox}}$	Neutrophils, macrophages, VSMCs, ECs, fibroblasts	5, 13, 24, 78, 84
$\text{p47}^{\text{phox}}$	Neutrophils, macrophages, VSMCs, ECs, fibroblasts	1, 4, 13, 64, 84
$\text{p41}^{\text{nox}}/\text{NOXO1}$	Colon	28
$\text{p67}^{\text{phox}}$	Neutrophils, macrophages	13, 54, 64, 84
$\text{p51}^{\text{nox}}/\text{NOXA-1}$	Colon	28
p40	Neutrophils, macrophages, VSMCs, ECs, fibroblasts	13, 54, 64, 84
rac	Neutrophils, macrophages, VSMCs, ECs	13, 54, 68

ECs, endothelial cells.

models of disease, as well as in human diseases. It is important to note that in cardiovascular diseases, not only the vascular oxidase, but also the phagocytic NADPH oxidase, plays an important role in  $\cdot\text{O}_2^-$  production, because monocytes and lymphocytes can infiltrate cardiovascular tissues (73).

Several works have established the role of the vascular NADPH oxidases in the development and progression of hypertension in numerous animals. Chronic infusion of angiotensin II (Ang II) in rats resulted in hypertensive animals, in correlation with an increased NADPH oxidase-derived  $\cdot\text{O}_2^-$  generation and up-regulated *p22<sup>phox</sup>* mRNA levels (25). Enhanced  $\cdot\text{O}_2^-$  production present in the aorta of DOCA-salt hypertensive rats is associated with increased NADPH oxidase activity and *p22<sup>phox</sup>* expression (7). Finally, we reported an enhanced NADPH oxidase-driven  $\cdot\text{O}_2^-$  production associated with an up-regulated *p22<sup>phox</sup>* mRNA expression in the aorta of adult spontaneously hypertensive rats (SHR), which present endothelial dysfunction and vascular wall hypertrophy (91). In the same work, we demonstrated that activated NADPH oxidase in the aorta of SHR depended on humoral and hemodynamic factors.

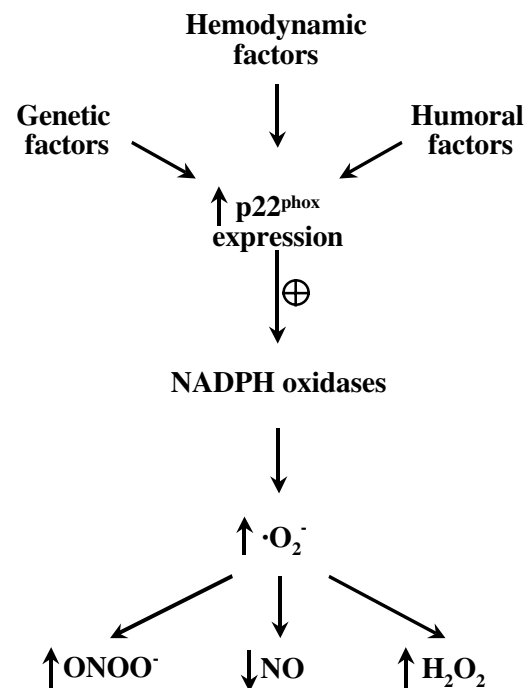
Although there is little information concerning the role of NADPH oxidases in human hypertension, some studies have demonstrated its implication. Recently, Ang II-induced  $\cdot\text{O}_2^-$  production has been described to be increased in vascular smooth muscle cells (VSMCs) isolated from hypertensive patients (83). Pettit *et al.* have shown that the NADPH oxidase-dependent  $\cdot\text{O}_2^-$  production is increased in lymphoblasts derived from hypertensive subjects (69). Furthermore, we have described the increased  $\cdot\text{O}_2^-$  production, dependent on NADPH oxidase, in stimulated mononuclear cells extracted from hypertensive patients (23).

Among the different components of the NADPH oxidase systems, *p22<sup>phox</sup>* emerges as an essential subunit for NADPH oxidase activity. Ushio-Fukai *et al.* demonstrated that the use of *p22<sup>phox</sup>* antisense technology prevented the increased NADPH oxidase-mediated  $\cdot\text{O}_2^-$  production induced by Ang II (86). Interestingly, many of the stimuli that activate vascular and phagocytic NADPH oxidases also increase *p22<sup>phox</sup>* expression. Among humoral factors, Ang II, TNF- $\alpha$ , interferon- $\gamma$ , and glucose are noteworthy (12, 17, 22, 25). Hemodynamic factors such as cyclic strain are also able to increase *p22<sup>phox</sup>* expression (59).

In summary, these studies suggest that *p22<sup>phox</sup>* overexpression may be a critical determinant of NADPH oxidase activation. It is important to note that, in addition to humoral and hemodynamic factors, the genetic background may also play an important role in the regulation of the NADPH oxidase systems (Fig. 1).

### *p22<sup>phox</sup>* GENE POLYMORPHISMS

During the past decade, the identification of NADPH oxidases as the major source of phagocytic and vascular  $\cdot\text{O}_2^-$  in cardiovascular diseases has provoked a significant interest in the possible association of polymorphisms in genes encoding NADPH oxidase subunits with cardiovascular disease susceptibility. A particular interest has been shown for the gene encoding the *p22<sup>phox</sup>* subunit, which possesses a significant

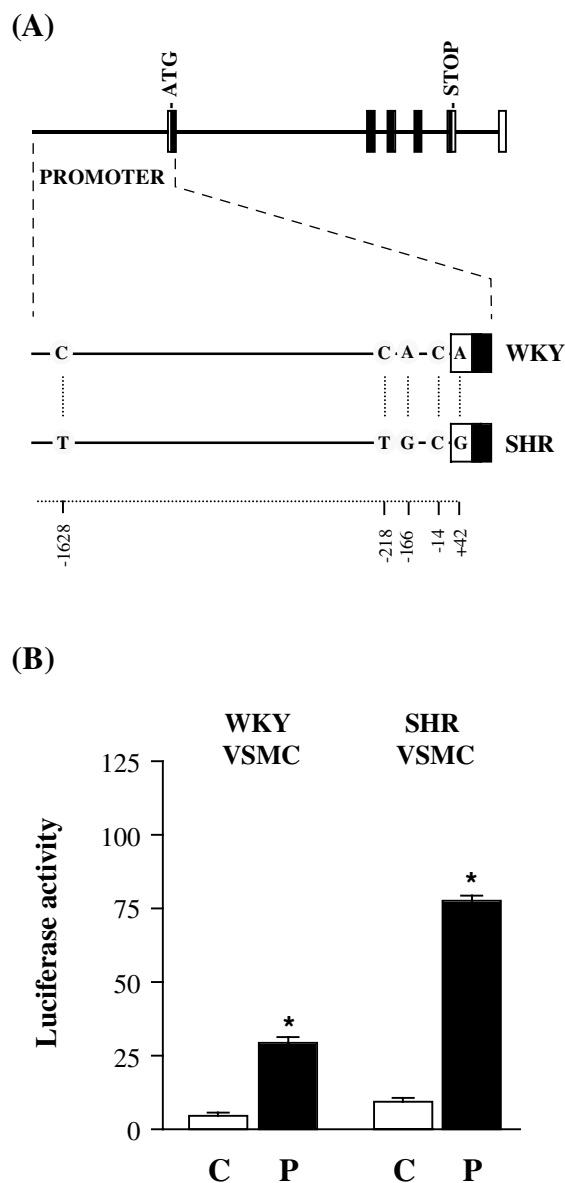


**FIG. 1. NADPH oxidase activation and functional consequences in arterial hypertension.** Besides multiple humoral agonists and hemodynamic forces, genetic changes may be involved in the activation of the NADPH oxidase systems by modulating the expression of NADPH oxidase subunits, including the *p22<sup>phox</sup>* protein. An enhanced  $\cdot\text{O}_2^-$  anion production driven by NADPH oxidase activation is involved in endothelial dysfunction by decreasing  $\cdot\text{NO}$  bioavailability and by increasing  $\text{ONOO}^-$  generation, and in media hypertrophy through the accumulation of  $\text{H}_2\text{O}_2$ .

number of genetic polymorphisms within the promoter and coding sequence, some of which are able to influence gene expression and NADPH oxidase activity.

### *p22<sup>phox</sup>* gene in genetic hypertension

Among the different studies performed in experimental hypertension, our group has demonstrated that increased *p22<sup>phox</sup>* expression correlated with activation of NADPH oxidase enzyme in the aorta of SHR (91). As SHR is a genetic model of hypertension, up-regulation of the oxidase *p22<sup>phox</sup>* subunit in SHR might be a consequence of alterations in the sequence of the *p22<sup>phox</sup>* gene (93). In line with this hypothesis, we characterized the 5'-end of the rat *p22<sup>phox</sup>* gene and identified five polymorphisms in this region of the SHR *p22<sup>phox</sup>* gene, one polymorphism located in the nontranslated region of exon 1 (+42), and four polymorphisms situated within the promoter region (−14, −166, −218, and −1,628). Interestingly, the higher promoter activity exhibited by the polymorphic promoter of the SHR *p22<sup>phox</sup>* gene evidenced a functional role for these allelic variants, thus suggesting that these polymorphisms might be involved in overexpression of the *p22<sup>phox</sup>* gene in the vascular wall of the SHR (92) (Fig. 2).



**FIG. 2. Influence of gene polymorphisms on *p22<sup>phox</sup>* gene promoter activity.** (A) Identification of the polymorphic variants in WKY and SHR *p22<sup>phox</sup>* gene promoters. ATG represents the translation initiation codon. (B) Transfection experiments with the SHR polymorphic promoter (P) and the WKY control promoter (C) into VSMCs from WKY and SHR. Histograms express relative luciferase activity of the *p22<sup>phox</sup>* promoter. \* $p < 0.05$  compared with WKY control promoter (adapted from reference 92).

### CYBA gene in human hypertension

*CYBA*, the human gene that encodes the *p22<sup>phox</sup>* protein, is situated on chromosome 16q24 (18, 67), and possesses several polymorphic regions both in the promoter and in the coding sequence (14). Some of these polymorphic sites (Fig. 3) have been associated with a number of cardiovascular diseases, such as hypertension, coronary artery disease (CAD), and myocardial infarction, all of which associate with an im-

paired endothelium-dependent vasodilation (45). It has been suggested that polymorphisms in the *CYBA* gene reduced the ability of *p22<sup>phox</sup>* subunit to anchor *gp91<sup>phox</sup>*, thus conditioning the stability of the b558 cytochrome and altering the NADPH oxidase activity (35).

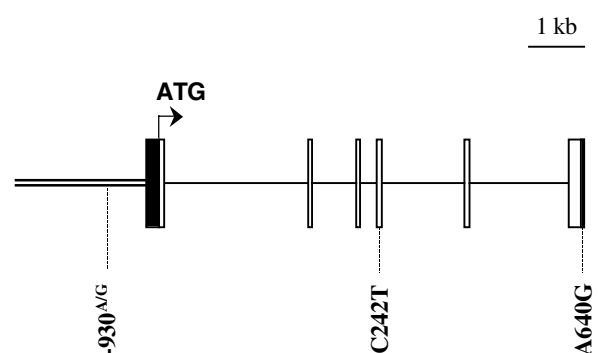
**A640G polymorphism.** The A640G gene polymorphism is located in the 3'-untranslated region of *CYBA* with no frank amino acid substitution (15, 71). It has been suggested that this genetic variant might modify mRNA processing and stability, thus influencing *p22<sup>phox</sup>* biosynthesis.

Gardemann *et al.* showed that the association of the A640G polymorphism with the presence and extent of CAD was stronger in hypertensives than in normotensives (26). Thus, a role for *CYBA* polymorphisms in NADPH oxidase activity can be hypothesized, contributing to the development of atherosclerosis in essential hypertension.

**C242T polymorphism.** The C242T polymorphism is located in exon 4, at position 242, of *CYBA* (18). It generates a CAC-TAC codon change, which results in a nonconservative substitution of histidine-72 by a tyrosine residue in the heme-binding site of the *p22<sup>phox</sup>* protein. It has been suggested that this replacement leads to a loss of oxidative function and to a reduced production of ROS and oxidative stress in the vasculature (88).

The association of the *CYBA* C242T polymorphism with atherosclerosis has been studied previously, although the results have been conflicting. Inoue *et al.* found that the T allele conferred protection against atherosclerosis in a Japanese population (37). However, other studies either did not find any association of the C242T polymorphism with atherosclerotic disease or even showed the opposite effect, that the T allele is significantly associated with progression of atherosclerosis (9) and cerebrovascular disease (39).

In the past decade, a functional role for C242T polymorphism in NADPH oxidase activity in physiological and pathophysiological conditions has been suggested. Guzik *et al.* demonstrated that 242T allele is associated with reduced vas-



**FIG. 3. Identification of the *CYBA* polymorphisms associated with cardiovascular diseases.** Boxes represent exons, single lines indicate introns, and the double line corresponds to the promoter region. An arrow points to the translation initiation ATG codon in exon 1. Black and white boxes indicate the nontranslated and the translated regions of exons, respectively.

cular NADPH oxidase activity in saphenous veins of CAD patients, independently of other clinical risk factors (32). More recently, Wyche *et al.* have shown that the NADPH oxidase-dependent phagocytic respiratory burst activity in homozygous healthy individuals with the T allele is significantly lower than that of wild-type carriers and heterozygous healthy individuals (89).

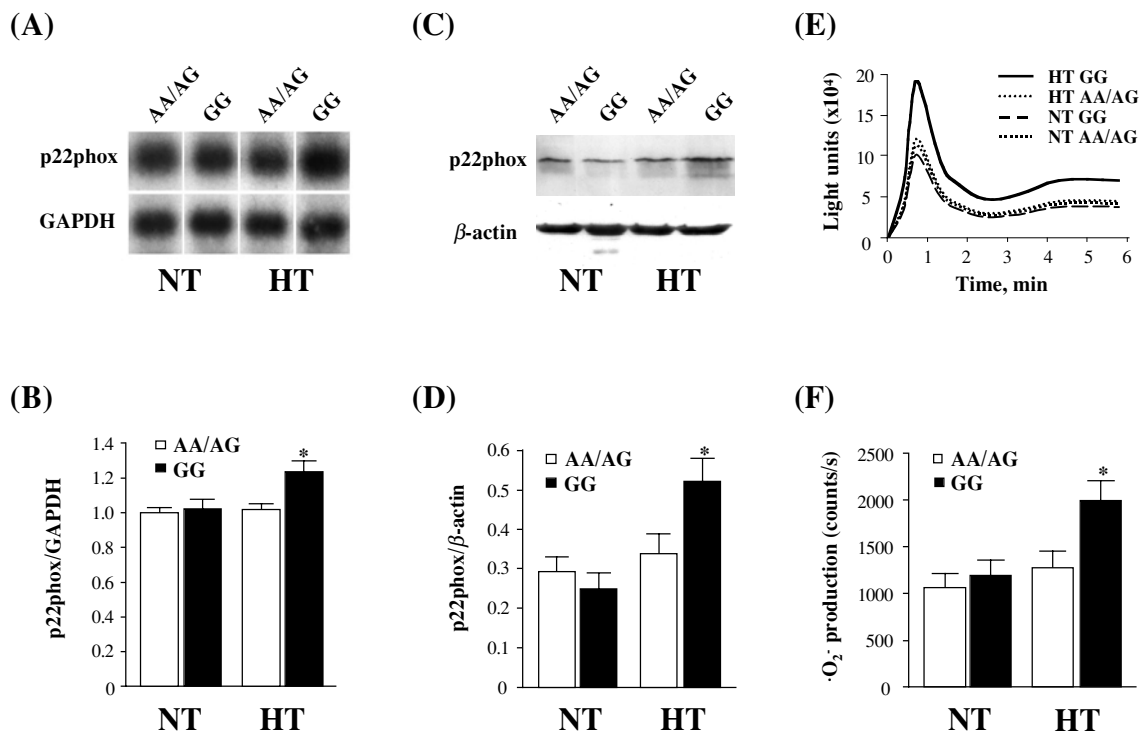
In agreement with these data, further studies have elucidated a significant impact of the C242T polymorphism on oxidative stress and endothelial function. Recently, Stanger *et al.* found no significant difference among genotypes in serum concentrations of malondialdehyde, an end product of lipid peroxidation (79). In contrast, Nakano *et al.* have demonstrated a diminished circulating concentration of oxidized high-density lipoprotein in CT/TT genotypes compared with CC genotypes in type 2 diabetic patients (63). Moreover, Schächinger *et al.* showed that carriers of the CC genotype revealed a significantly blunted endothelium-dependent dilator response, which was independent of other risk factors or atherosclerosis (76). Collectively, the association of the C242T polymorphism with NADPH oxidase activity and endothelial function implies this genetic variant might be a relevant clinical marker of CAD.

On the other hand, Raijmakers *et al.* reported a lack of association between the C242T polymorphism and preeclampsia (72).

Nevertheless, in a recent study, we have found a significant association of the C242T polymorphism with essential hypertension in a Spanish population (95). Moreover, we have found that phagocytic NADPH oxidase activity was significantly higher in CC than in TC/TT hypertensives, whereas no differences were found between genotypes in the normotensive control population, suggesting that the effects of C242T polymorphism may become evident in the presence of cardiovascular risk factors.

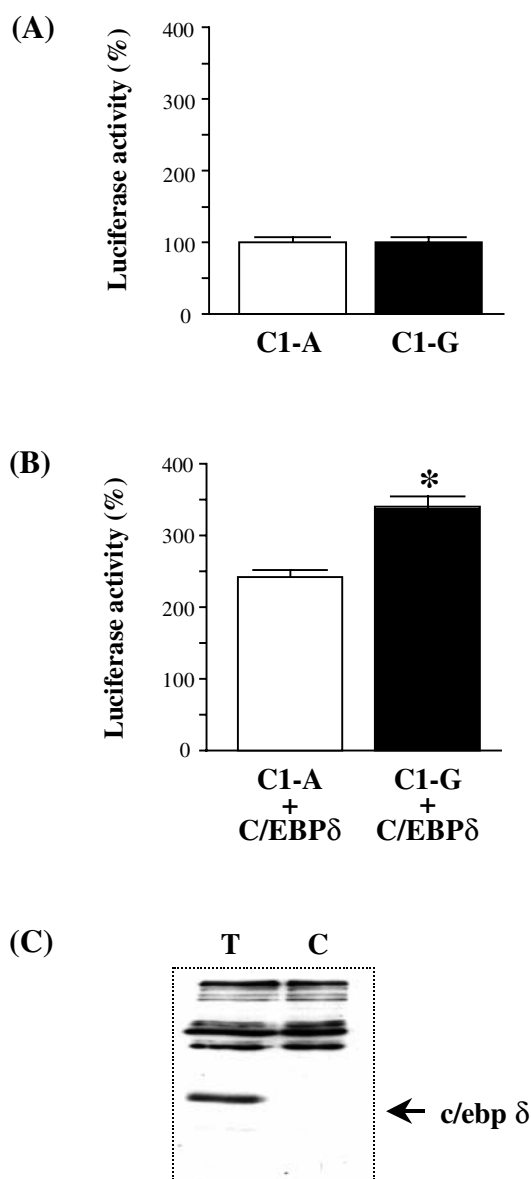
**−930<sup>A/G</sup> polymorphism.** The −930<sup>A/G</sup> polymorphism is located in the promoter region of *CYBA*, and it has been speculated to have a functional effect on the modulation of *CYBA* transcriptional activity.

The association of the *CYBA* −930<sup>A/G</sup> polymorphism with hypertension has been studied recently. Moreno *et al.* found that the −930<sup>A/G</sup> polymorphism was associated with hypertension in a Spanish population (61). Frequencies of the GG, GA, and AA genotypes were 0.34, 0.43, and 0.23 for the normotensive group and 0.41, 0.47, and 0.12 for the hypertensive group. In line with these results, San José *et al.* have recently demonstrated that the −930<sup>GG</sup> genotype is associated with increased *p22<sup>phox</sup>* expression and NADPH oxidase activity in phagocytic cells of hypertensive patients (Fig. 4) (75). In contrast, no dif-



**FIG. 4.** Effect of the −930<sup>A/G</sup> polymorphism on *p22<sup>phox</sup>* expression and NADPH oxidase activity. (A) The expression of phagocytic *p22<sup>phox</sup>* mRNA was adjusted for the expression of the housekeeping gene glyceraldehyde-3-phosphate dehydrogenase (GAPDH). (B) Estimation of the *p22<sup>phox</sup>* mRNA expression. \**p* < 0.05 compared with AA/AG hypertensives (HT) and with AA/AG and GG normotensives (NT). (C) The expression of phagocytic *p22<sup>phox</sup>* protein was adjusted for the expression of the housekeeping gene β-actin. (D) Estimation of the *p22<sup>phox</sup>* protein expression. \**p* < 0.05 compared with AA/AG hypertensives and with AA/AG and GG normotensives. (E) ·O<sub>2</sub><sup>−</sup> production was determined by lucigenin chemiluminescence after phorbol myristate acetate stimulation. (F) Estimation of the NADPH oxidase activity. \**p* < 0.05 compared with AA/AG hypertensives and with AA/AG and GG normotensives (adapted from reference 75).





**FIG. 5. Reporter gene expression assays with *CYBA* promoter allelic constructs.** (A) Changes in luciferase levels induced by the A-to-G substitution at the  $-930$  polymorphic site in A7r5 cells. C1-A, A allelic construct; C1-G, G allelic construct. (B) Effect of C/EBP $\delta$  cotransfection on luciferase levels of p22<sup>phox</sup> promoter allelic constructs in A7r5 cells. \* $p < 0.05$ . (C) Western blot experiment shows a positive immunodetection of C/EBP $\delta$  protein in C/EBP $\delta$ -transfected A7r5 cells. C, control A7r5 cells; T, C/EBP $\delta$ -transfected A7r5 cells. An arrow points to the expressed protein band of 35 kDa (adapted from reference 75).

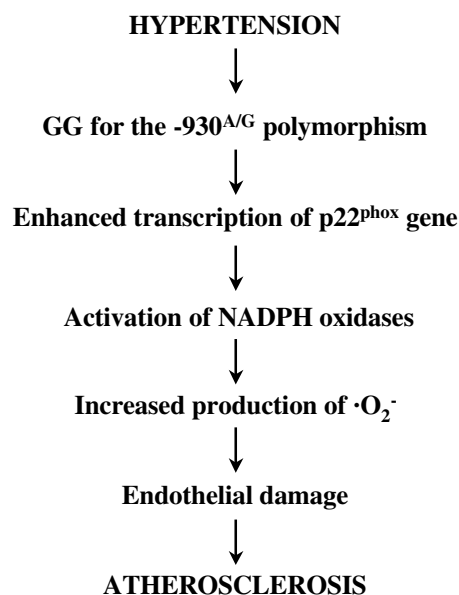
ferences were found in the NADPH oxidase expression and activity between genotypes within the normotensive group. In accordance with these *in vivo* measurements, *in vitro* experiments showed that the A-to-G substitution results in an increased *CYBA* gene promoter activity in hypertensive, but not in normotensive, VSMCs. In agreement with this, Wyche *et al.* have shown no differences in NADPH oxidase-dependent

phagocytic respiratory burst activity between  $-930^{A/G}$  polymorphism genotypes in healthy young individuals (89). Collectively, these data suggest that the  $-930^{A/G}$  polymorphism may be functionally relevant in the control of the *CYBA* gene expression under hypertensive conditions.

Although the mechanisms that underlie this control remain unclear, one possibility is that the presence of the G allele modulates the transcription of the p22<sup>phox</sup> gene. In this regard, an analysis of the promoter sequence shows that the  $-930$  polymorphic site lies on a potential binding site for CCAAT enhancer-binding protein (C/EBP) transcription factors. Furthermore, our results showing the effect of C/EBP $\delta$  overexpression on higher transcriptional activity of the G than of the A allelic p22<sup>phox</sup> promoter construct allow us to suggest the involvement of C/EBPs in the hypertensive phenotype (Fig. 5). Several findings that underlie the role of C/EBPs in hypertension support this possibility. C/EBP $\delta$  expression is nearly absent in Wistar Kyoto rat (WKY) VSMCs, whereas it is abnormally increased in SHR VSMCs (44). Changes in angiotensinogen mRNA expression associated with the  $-217^{A/G}$  polymorphism of this gene are regulated by several members of the C/EBP family (40). Inflammatory cytokines up-regulate C/EBP expression (70), and cytokine levels have been reported to be increased in phagocytic cells in hypertensives (19).

Alternatively, the G allele may be in linkage disequilibrium with other genetic variants that could influence the transcriptional activity of *CYBA*. In this regard, we have identified other genetic variants in the p22<sup>phox</sup> promoter. One of them, the  $-675^{A/T}$  polymorphism, is associated with hypertension, although its functional implication in NADPH oxidase activity remains to be elucidated (62).

The impact of the  $-930^{A/G}$  polymorphism on oxidative stress and endothelial function is currently being elucidated. Recent research demonstrated the involvement of the NADPH oxidases in the uncoupling of the endothelial  $\cdot\text{NO}$  synthase,



**FIG. 6. The  $-930^{A/G}$  polymorphism as a genetic marker of oxidative stress in hypertension.**

which favors the diminished  $\cdot\text{NO}$  generation in hypertensive animals (51). Previous studies reported reduced levels of  $\cdot\text{NO}$  metabolites (nitrate and nitrite) in the hypertensive population (57, 85). Interestingly, the higher  $\cdot\text{O}_2^-$  generation observed in  $-930^{\text{GG}}$  hypertensives associated with a diminished production of systemic  $\cdot\text{NO}$ . Finally, GG hypertensives associated with higher plasma levels of von Willebrand factor (94). As increased levels of von Willebrand factor have been found to be associated with the extent of damage in the vascular endothelium and increased subsequent occurrence of atherosclerotic cardiovascular events (8, 56, 58, 60), these findings suggest that the  $-930^{\text{A/G}}$  polymorphism may be a novel genetic marker of endothelial damage and of risk of atherosclerosis in essential hypertension (Fig. 6).

## CONCLUSION AND PERSPECTIVES

Increased  $\cdot\text{O}_2^-$  generation is an important feature of the vascular phenotype in human hypertension, as it affects  $\cdot\text{NO}$  bioavailability,  $\text{ONOO}^-$  generation, and redox-sensitive cell-signaling pathways. The NADPH oxidases are important sources of  $\cdot\text{O}_2^-$  in human vessels. Genetic factors, including polymorphic variants in the genes encoding the NADPH oxidase subunits, modulate  $\cdot\text{O}_2^-$  production, influencing hypertension and the progression of cardiovascular disease. The identification of functional polymorphisms in *CYBA*, associated with a higher NADPH oxidase-dependent  $\cdot\text{O}_2^-$  production, could represent another criterion to bear in mind for the adequate characterization of patients with a genetically increased susceptibility to oxidative stress, which would constitute an appropriate population for therapeutic interventions with antioxidants.

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## ABBREVIATIONS

Ang II, angiotensin II; CAD, coronary artery disease; C/EBP, CCAAT enhancer-binding protein;  $\text{H}_2\text{O}_2$ , hydrogen peroxide;  $\cdot\text{NO}$ , nitric oxide;  $\cdot\text{O}_2^-$ , superoxide anion;  $\text{ONOO}^-$ , peroxynitrite; ROS, reactive oxygen species; SHR, spontaneously hypertensive rats;  $\text{TNF-}\alpha$ , tumor necrosis factor- $\alpha$ ; VSMCs, vascular smooth muscle cells; WKY, Wistar Kyoto rats.

## REFERENCES

- Al-Mehdi AB, Zhao G, Dodia C, Tozawa K, Costa K, Muzykantov V, Ross C, Blecha F, Dinauer M, and Fisher AB. Endothelial NADPH oxidase as the source of oxidants in lungs exposed to ischemia or high  $\text{K}^+$ . *Circ Res* 83: 730–737, 1998.
- Banfi B, Molnar G, Maturana A, Steger K, Hegedus B, De-maurex N, and Krause KH. A  $\text{Ca}^{2+}$ -activated NADPH oxidase in testis, spleen, and lymph nodes. *J Biol Chem* 276: 37594–37601, 2001.
- Banfi B, Malgrange B, Knisz J, Steger K, Dubois-Dauphin M, and Krause KH. NOX3, a superoxide-generating NADPH oxidase of the inner ear. *J Biol Chem* 279: 46065–46072, 2004.
- Barry-Lane PA, Patterson C, van der Merwe M, Hu Z, Holland SM, Yeh ET, and Runge MS. p47phox is required for atherosclerotic lesion progression in ApoE(–/–) mice. *J Clin Invest* 108: 1513–1522, 2001.
- Bayraktutan U, Blayney L, and Shah AM. Molecular characterization and localization of the NAD(P)H oxidase components gp91-phox and p22-phox in endothelial cells. *Arterioscler Thromb Vasc Biol* 20: 1903–1911, 2000.
- Beckman JS and Koppenol WH. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. *Am J Physiol* 271: C1424–C1437, 1996.
- Beswick RA, Dorrance AM, Leite R, and Webb RC. NADH/NADPH oxidase and enhanced superoxide production in the mineralocorticoid hypertensive rat. *Hypertension* 38: 1107–1111, 2001.
- Blann AD, Naqvi T, Waite M, and McCollum CN. von Willebrand factor and endothelial damage in essential hypertension. *J Hum Hypertens* 7: 107–111, 1993.
- Cahilly C, Ballantyne CM, Lim DS, Gotto A, and Marian AJ. A variant of p22(phox), involved in generation of reactive oxygen species in the vessel wall, is associated with progression of coronary atherosclerosis. *Circ Res* 86: 391–395, 2000.
- Cai H and Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. *Circ Res* 87: 840–844, 2000.
- Cheng G, Cao Z, Xu X, van Meir EG, and Lambeth JD. Homologs of gp91phox: cloning and tissue expression of Nox3, Nox4, and Nox5. *Gene* 269: 131–140, 2001.
- Cosentino F, Eto M, De Paolis P, van der Loo B, Bachschmid M, Ullrich V, Kouroedov A, Delli Gatti C, Joch H, Volpe M, and Luscher TF. High glucose causes upregulation of cyclooxygenase-2 and alters prostanoid profile in human endothelial cells: role of protein kinase C and reactive oxygen species. *Circulation* 107: 1017–1023, 2003.
- Cross AR and Segal AW. The NADPH oxidase of professional phagocytes—prototype of the NOX electron transport chain systems. *Biochim Biophys Acta* 1657: 1–22, 2004.
- Cross AR, Noack D, Rae J, Curnutte JT, and Heyworth PG. Hematologically important mutations: the autosomal recessive forms of chronic granulomatous disease (first update). *Blood Cells Mol Dis* 26: 561–565, 2000.
- de Boer M, de Klein A, Hossle JP, Seger R, Corbeel L, Weening RS, and Roos D. Cytochrome b558-negative, autosomal recessive chronic granulomatous disease: two new mutations in the cytochrome b558 light chain of the NADPH oxidase (p22-phox). *Am J Hum Genet* 51: 1127–1135, 1992.

16. De Deken X, Wang D, Many MC, Costagliola S, Libert F, Vassart G, Dumont JE, and Miot F. Cloning of two human thyroid cDNAs encoding new members of the NADPH oxidase family. *J Biol Chem* 275: 23227–23233, 2000.
17. De Keulenaer GW, Alexander RW, Ushio-Fukai M, Ishizaka N, and Griendling KK. Tumour necrosis factor alpha activates a p22phox-based NADH oxidase in vascular smooth muscle. *Biochem J* 329: 653–657, 1998.
18. Dinanier MC, Pierce EA, Bruns GA, Curnutte JT, and Orkin SH. Human neutrophil cytochrome b light chain (p22-phox). Gene structure, chromosomal location, and mutations in cytochrome-negative autosomal recessive chronic granulomatous disease. *J Clin Invest* 86: 1729–1737, 1990.
19. Dorffle Y, Latsch C, Stuhlmüller B, Schreiber S, Scholze S, Burmester GR, and Scholze J. Preactivated peripheral blood monocytes in patients with essential hypertension. *Hypertension* 34: 113–117, 1999.
20. Droge W. Free radicals in the physiological control of cell function. *Physiol Rev* 82: 47–95, 2002.
21. Dupuy C, Ohayon R, Valent A, Noel-Hudson MS, Deme D, and Virion A. Purification of a novel flavoprotein involved in the thyroid NADPH oxidase. Cloning of the porcine and human cDNAs. *J Biol Chem* 274: 37265–37269, 1999.
22. Dusi S, Donini M, Lissandrini D, Mazzi P, Bianca VD, and Rossi F. Mechanisms of expression of NADPH oxidase components in human cultured monocytes: role of cytokines and transcriptional regulators involved. *Eur J Immunol* 31: 929–938, 2001.
23. Fortuno A, Oliván S, Belóqui O, San Jose G, Moreno MU, Díez J, and Zalba G. Association of increased phagocytic NADPH oxidase-dependent superoxide production with diminished nitric oxide generation in essential hypertension. *J Hypertens* 22: 2169–2175, 2004.
24. Fukui T, Lassegue B, Kai H, Alexander RW, and Griendling KK. Cytochrome b-558 alpha-subunit cloning and expression in rat aortic smooth muscle cells. *Biochim Biophys Acta* 1231: 215–219, 1995.
25. Fukui T, Ishizaka N, Rajagopalan S, Laursen JB, Capers Q 4th, Taylor WR, Harrison DG, de Leon H, Wilcox JN, and Griendling KK. p22phox mRNA expression and NADPH oxidase activity are increased in aortas from hypertensive rats. *Circ Res* 80: 45–51, 1997.
26. Gardemann A, Mages P, Katz N, Tillmanns H, and Haberbosch W. The p22 phox A640G gene polymorphism but not the C242T gene variation is associated with coronary heart disease in younger individuals. *Atherosclerosis* 145: 315–323, 1999.
27. Geiszt M, Kopp JB, Várnai P, and Leto TL. Identification of renox, an NAD(P)H oxidase in kidney. *Proc Natl Acad Sci USA* 97: 8010–8014, 2000.
28. Geiszt M, Lekstrom K, Witta J, and Leto TL. Proteins homologous to p47phox and p67phox support superoxide production by NAD(P)H oxidase 1 in colon epithelial cells. *J Biol Chem* 278: 20006–20012, 2003.
29. Griendling KK and FitzGerald GA. Oxidative stress and cardiovascular injury: Part II: Animal and human studies. *Circulation* 108: 2034–2040, 2003.
30. Griendling KK and Harrison DG. Dual role of reactive oxygen species in vascular growth. *Circ Res* 85: 562–563, 1999.
31. Griendling KK, Sorescu D, and Ushio-Fukai M. NAD(P)H oxidase: role in cardiovascular biology and disease. *Circ Res* 86: 494–501, 2000.
32. Guzik TJ, West NE, Black E, McDonald D, Ratnatunga C, Pillai R, and Channon KM. Functional effect of the C242T polymorphism in the NAD(P)H oxidase p22phox gene on vascular superoxide production in atherosclerosis. *Circulation* 102: 1744–1747, 2000.
33. Guzik TJ, Mussa S, Gastaldi D, Sadowski J, Ratnatunga C, Pillai R, and Channon KM. Mechanisms of increased vascular superoxide production in human diabetes mellitus: role of NAD(P)H oxidase and endothelial nitric oxide synthase. *Circulation* 105: 1656–1662, 2002.
34. Harrison D, Griendling KK, Landmesser U, Hornig B, and Drexler H. Role of oxidative stress in atherosclerosis. *Am J Cardiol* 91: 7A–11A, 2003.
35. Huang J, Hitt ND, and Kleinberg ME. Stoichiometry of p22-phox and gp91-phox in phagocyte cytochrome b558. *Biochemistry* 34: 16753–16757, 1995.
36. Inoguchi T, Tsubouchi H, Etoh T, Kakimoto M, Sonta T, Utsumi H, Sumimoto H, Yu HY, Sonoda N, Inuo M, Sato N, Sekiguchi N, Kobayashi K, and Nawata H. A possible target of antioxidative therapy for diabetic vascular complications—vascular NAD(P)H oxidase. *Curr Med Chem* 10: 1759–1764, 2003.
37. Inoue N, Kawashima S, Kanazawa K, Yamada S, Akita H, and Yokoyama M. Polymorphism of the NADH/NADPH oxidase p22 phox gene in patients with coronary artery disease. *Circulation* 97: 135–137, 1998.
38. Irani K. Oxidant signaling in vascular cell growth, death, and survival: a review of the roles of reactive oxygen species in smooth muscle and endothelial cell mitogenic and apoptotic signaling. *Circ Res* 87: 179–183, 2000.
39. Ito D, Murata M, Watanabe K, Yoshida T, Saito I, Tanahashi N, and Fukuuchi Y. C242T polymorphism of NADPH oxidase p22 PHOX gene and ischemic cerebrovascular disease in the Japanese population. *Stroke* 31: 936–939, 2000.
40. Jain S, Tang X, Narayanan CS, Agarwal Y, Peterson SM, Brown CD, Ott J, and Kumar A. Angiotensinogen gene polymorphism at –217 affects basal promoter activity and is associated with hypertension in African-Americans. *J Biol Chem* 277: 36889–36896, 2002.
41. Kalinina N, Agrotis A, Tararak E, Antropova Y, Kanellakis P, Ilyinskaya O, Quinn MT, Smirnov V, and Bobik A. Cytochrome b558-dependent NAD(P)H oxidase-phox units in smooth muscle and macrophages of atherosclerotic lesions. *Arterioscler Thromb Vasc Biol* 22: 2037–2043, 2002.
42. Katsuyama M, Fan C, and Yabe-Nishimura C. NADPH oxidase is involved in prostaglandin F2alpha-induced hypertrophy of vascular smooth muscle cells: induction of NOX1 by PGF2alpha. *J Biol Chem* 277: 13438–13442, 2002.
43. Kikuchi H, Hikage M, Miyashita H, and Fukumoto M. NADPH oxidase subunit, gp91(phox) homologue, preferentially expressed in human colon epithelial cells. *Gene* 254: 237–243, 2000.
44. Kitami Y, Fukuoka T, Hiwada K, and Inagami T. A high level of CCAAT-enhancer binding protein-delta expression is a major determinant for markedly elevated differential gene expression of the platelet-derived growth factor-



- alpha receptor in vascular smooth muscle cells of genetically hypertensive rats. *Circ Res* 84: 64–73, 1999.
45. Kojda G and Harrison D. Interactions between NO and reactive oxygen species: pathophysiological importance in atherosclerosis, hypertension, diabetes and heart failure. *Cardiovasc Res* 43: 562–571, 1999.
  46. Krijnen PA, Meischl C, Hack CE, Meijer CJ, Visser CA, Roos D, and Niessen HW. Increased Nox2 expression in human cardiomyocytes after acute myocardial infarction. *J Clin Pathol* 56: 194–199, 2003.
  47. Kumar KV and Das UN. Are free radicals involved in the pathobiology of human essential hypertension? *Free Radic Res Commun* 19: 59–66, 1993.
  48. Kunsch C and Medford RM. Oxidative stress as a regulator of gene expression in the vasculature. *Circ Res* 85: 753–766, 1999.
  49. Lacy F, Kailasam MT, O'Connor DT, Schmid-Schonbein GW, and Parmer RJ. Plasma hydrogen peroxide production in human essential hypertension: role of heredity, gender, and ethnicity. *Hypertension* 36: 878–884, 2000.
  50. Lambeth JD. Nox/Duox family of nicotinamide adenine dinucleotide (phosphate) oxidases. *Curr Opin Hematol* 9: 11–17, 2002.
  51. Landmesser U, Dikalov S, Price SR, McCann L, Fukai T, Holland SM, Mitch WE, and Harrison DG. Oxidation of tetrahydrobiopterin leads to uncoupling of endothelial cell nitric oxide synthase in hypertension. *J Clin Invest* 111: 1201–1209, 2003.
  52. Lassegue B and Clempus RE. Vascular NAD(P)H oxidases: specific features, expression, and regulation. *Am J Physiol Regul Integr Comp Physiol* 285: R277–R297, 2003.
  53. Lassegue B and Griendling KK. Reactive oxygen species in hypertension; an update. *Am J Hypertens* 17: 852–860, 2004.
  54. Li JM and Shah AM. Intracellular localization and pre-assembly of the NADPH oxidase complex in cultured endothelial cells. *J Biol Chem* 277: 19952–19960, 2002.
  55. Li PF, Dietz R, and von Harsdorf R. Differential effect of hydrogen peroxide and superoxide anion on apoptosis and proliferation of vascular smooth muscle cells. *Circulation* 96: 3602–3609, 1997.
  56. Lip GY and Blann A. von Willebrand factor: a marker of endothelial dysfunction in vascular disorders? *Cardiovasc Res* 34: 255–265, 1997.
  57. Lyamina NP, Dolotovskaya PV, Lyamina SV, Malyshev IY, and Manukhina EB. Nitric oxide production and intensity of free radical processes in young men with high normal and hypertensive blood pressure. *Med Sci Monit* 9: CR304–CR310, 2003.
  58. Mannucci PM. von Willebrand factor: a marker of endothelial damage? *Arterioscler Thromb Vasc Biol* 18: 1359–1362, 1998.
  59. Matsushita H, Lee K, and Tsao PS. Cyclic strain induces reactive oxygen species production via an endothelial NAD(P)H oxidase. *J Cell Biochem* 81: 99–106, 2001.
  60. Morange PE, Simon C, Alessi MC, Luc G, Arveiler D, Ferrieres J, Amouyel P, Evans A, Ducimetiere P, and Juhan-Vague I. Endothelial cell markers and the risk of coronary heart disease: the Prospective Epidemiological Study of Myocardial Infarction (PRIME) study. *Circulation* 109: 1343–1348, 2004.
  61. Moreno MU, San Jose G, Orbe J, Paramo JA, Beloqui O, Diez J, and Zalba G. Preliminary characterisation of the promoter of the human p22(phox) gene: identification of a new polymorphism associated with hypertension. *FEBS Lett* 542: 27–31, 2003.
  62. Moreno MU, Fortuño A, San José G, Oliván S, Montoya A, Ros R, Beloqui O, Diez J, and Zalba G. New polymorphisms in the human p22phox gene: association studies with essential hypertension. *J Hypertens (Suppl 2)* 22: S6, 2004.
  63. Nakano T, Matsunaga S, Nagata A, and Maruyama T. NAD(P)H oxidase p22phox Gene C242T polymorphism and lipoprotein oxidation. *Clin Chim Acta* 335: 101–107, 2003.
  64. Pagano PJ, Clark JK, Cifuentes-Pagano ME, Clark SM, Callis GM, and Quinn MT. Localization of a constitutively active, phagocyte-like NADPH oxidase in rabbit aortic adventitia: enhancement by angiotensin II. *Proc Natl Acad Sci U S A* 94: 14483–14488, 1997.
  65. Paravicini TM and Sobey CG. Cerebral vascular effects of reactive oxygen species: recent evidence for a role of NADPH-oxidase. *Clin Exp Pharmacol Physiol* 30: 855–859, 2003.
  66. Park JB. Phagocytosis induces superoxide formation and apoptosis in macrophages. *Exp Mol Med* 35: 325–335, 2003.
  67. Parkos CA, Dinanier MC, Walker LE, Allen RA, Jesaitis AJ, and Orkin SH. Primary structure and unique expression of the 22-kilodalton light chain of human neutrophil cytochrome b. *Proc Natl Acad Sci U S A* 85: 3319–3323, 1988.
  68. Patterson C, Ruef J, Madamanchi NR, Barry-Lane P, Hu Z, Horaist C, Ballinger CA, Brasier AR, Bode C, and Runge MS. Stimulation of a vascular smooth muscle cell NAD(P)H oxidase by thrombin. Evidence that p47(phox) may participate in forming this oxidase in vitro and in vivo. *J Biol Chem* 274: 19814–19822, 1999.
  69. Pettit AI, Wong RK, Lee V, Jennings S, Quinn PA, and Ng LL. Increased free radical production in hypertension due to increased expression of the NADPH oxidase subunit p22(phox) in lymphoblast cell lines. *J Hypertens* 20: 677–683, 2002.
  70. Poli V. The role of C/EBP isoforms in the control of inflammatory and native immunity functions. *J Biol Chem* 273: 29279–29282, 1998.
  71. Rae J, Noack D, Heyworth PG, Ellis BA, Curnutte JT, and Cross AR. Molecular analysis of 9 new families with chronic granulomatous disease caused by mutations in CYBA, the gene encoding p22(phox). *Blood* 96: 1106–1112, 2000.
  72. Rajmakers MT, Roes EM, Steegers EA, and Peters WH. The C242T-polymorphism of the NADPH/NADH oxidase gene p22phox subunit is not associated with pre-eclampsia. *J Hum Hypertens* 16: 423–425, 2002.
  73. Rodriguez-Iturbe B, Zhan CD, Quiroz Y, Sindhu RK, and Vaziri ND. Antioxidant-rich diet relieves hypertension and reduces renal immune infiltration in spontaneously hypertensive rats. *Hypertension* 41: 341–346, 2003.

74. Salonen RM, Nyyssonen K, Kaikkonen J, Porkkala-Sarataho E, Voutilainen S, Rissanen TH, Tuomainen TP, Valkonen VP, Ristonmaa U, Lakka HM, Vanharanta M, Salonen JT, and Poulsen HE. Six-year effect of combined vitamin C and E supplementation on atherosclerotic progression: the Antioxidant Supplementation in Atherosclerosis Prevention (ASAP) Study. *Circulation* 107: 947–953, 2003.
75. San Jose G, Moreno MU, Olivan S, Beloqui O, Fortuno A, Diez J, and Zalba G. Functional effect of the p22phox –930A/G polymorphism on p22phox expression and NADPH oxidase activity in hypertension. *Hypertension* 44: 163–169, 2004.
76. Schächinger V, Britten MB, Dimmeler S, and Zeiher AM. NADH/NADPH oxidase p22 phox gene polymorphism is associated with improved coronary endothelial vasodilator function. *Eur Heart J* 22: 96–101, 2001.
77. Shiose A, Kuroda J, Tsuruya K, Hirai M, Hirakata H, Naito S, Hattori M, Sakaki Y, and Sumimoto H. A novel superoxide-producing NAD(P)H oxidase in kidney. *J Biol Chem* 276: 1417–1423, 2001.
78. Sorescu D, Weiss D, Lassegue B, Clempus RE, Szocs K, Sorescu GP, Valppu L, Quinn MT, Lambeth JD, Vega JD, Taylor WR, and Griendling KK. Superoxide production and expression of Nox family proteins in human atherosclerosis. *Circulation* 105: 1429–1435, 2002.
79. Stanger O, Renner W, Khoschsorur G, Rigler B, and Wascher TC. NADH/NADPH oxidase p22 phox C242T polymorphism and lipid peroxidation in coronary artery disease. *Clin Physiol* 21: 718–722, 2001.
80. Stephens NG, Parsons A, Schofield PM, Kelly F, Cheeseman K, and Mitchinson MJ. Randomised controlled trial of vitamin E in patients with coronary disease: Cambridge Heart Antioxidant Study (CHAOS). *Lancet* 347: 781–786, 1996.
81. Taniyama Y and Griendling KK. Reactive oxygen species in the vasculature: molecular and cellular mechanisms. *Hypertension* 42: 1075–1081, 2003.
82. Touyz RM. Reactive oxygen species as mediators of calcium signaling by angiotensin II: implications in vascular physiology and pathophysiology. *Antioxid Redox Signal* 7: 1302–1314, 2005.
83. Touyz RM and Schiffrin EL. Increased generation of superoxide by angiotensin II in smooth muscle cells from resistance arteries of hypertensive patients: role of phospholipase D-dependent NAD(P)H oxidase-sensitive pathways. *J Hypertens* 19: 1245–1254, 2001.
84. Touyz RM, Chen X, Tabet F, Yao G, He G, Quinn MT, Pagano PJ, and Schiffrin EL. Expression of a functionally active gp91phox-containing neutrophil-type NAD(P)H oxidase in smooth muscle cells from human resistance arteries: regulation by angiotensin II. *Circ Res* 90: 1205–1213, 2002.
85. Turi S, Friedman A, Bereczki C, Papp F, Kovacs J, Karg E, and Nemeth I. Oxidative stress in juvenile essential hypertension. *J Hypertens* 21: 145–152, 2003.
86. Ushio-Fukai M, Zafari AM, Fukui T, Ishizaka N, and Griendling KK. p22phox is a critical component of the superoxide-generating NADH/NADPH oxidase system and regulates angiotensin II-induced hypertrophy in vascular smooth muscle cells. *J Biol Chem* 271: 23317–23321, 1996.
87. Wang HD, Xu S, Johns DG, Du Y, Quinn MT, Cayatte AJ, and Cohen RA. Role of NADPH oxidase in the vascular hypertrophic and oxidative stress response to angiotensin II in mice. *Circ Res* 88: 947–953, 2001.
88. Whitehead AS and FitzGerald GA. Twenty-first century phox: not yet ready for widespread screening. *Circulation* 103: 7–9, 2001.
89. Wyche KE, Wang SS, Griendling KK, Dikalov SI, Austin H, Rao S, Fink B, Harrison DG, and Zafari AM. C242T CYBA polymorphism of the NADPH oxidase is associated with reduced respiratory burst in human neutrophils. *Hypertension* 43: 1246–1251, 2004.
90. Yang S, Madyastha P, Bingel S, Ries W, and Key L. A new superoxide-generating oxidase in murine osteoclasts. *J Biol Chem* 276: 5452–5458, 2001.
91. Zalba G, Beaumont FJ, San Jose G, Fortuno A, Fortuno MA, Etayo JC, and Diez J. Vascular NADH/NADPH oxidase is involved in enhanced superoxide production in spontaneously hypertensive rats. *Hypertension* 35: 1055–1061, 2000.
92. Zalba G, San Jose G, Beaumont FJ, Fortuno MA, Fortuno A, and Diez J. Polymorphisms and promoter overactivity of the p22(phox) gene in vascular smooth muscle cells from spontaneously hypertensive rats. *Circ Res* 88: 217–222, 2001.
93. Zalba G, San Jose G, Moreno MU, Fortuno MA, Fortuno A, Beaumont FJ, and Diez J. Oxidative stress in arterial hypertension: role of NAD(P)H oxidase. *Hypertension* 38: 1395–1399, 2001.
94. Zalba G, Moreno MU, San Jose G, Beloqui O, Ros R, Fortuno A, and Diez J. The –930A/G polymorphism in the p22phox gene possesses functional effect on superoxide production in essential hypertension. *Circulation (Suppl IV)* 108: 46, 2003.
95. Zalba G, San José G, Moreno MU, Oliván S, Ros R, Montoya A, Beloqui O, Fortuño A, and Díez J. Functional effect of the C242T polymorphism in the NAD(P)H oxidase p22phox gene on superoxide production in essential hypertension. *J Hypertens (Suppl 2)* 22: S346, 2004.

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2. Augusto C. Montezano, Rhian M. Touyz. 2012. Oxidative stress, Noxs, and hypertension: Experimental evidence and clinical controversies. *Annals of Medicine* **44**:S1, S2-S16. [[CrossRef](#)]
3. R. Schreiber, M. C. Ferreira-Sae, A. C. Tucunduva, J. G. Mill, Felipe O. Costa, J. E. Krieger, K. G. Franchini, A. C. Pereira, W. Nadruz. 2012. CYBA C242T polymorphism is associated with obesity and diabetes mellitus in Brazilian hypertensive patients. *Diabetic Medicine* no-no. [[CrossRef](#)]
4. Ashwani K Khanna, Jianping Xu, Mandeep R Mehra. 2011. Antioxidant N-acetyl cysteine reverses cigarette smoke-induced myocardial infarction by inhibiting inflammation and oxidative stress in a rat model. *Laboratory Investigation* . [[CrossRef](#)]
5. Javier Piérola, Alexandra Alemany, Aina Yañez, Mónica de-la-Peña, Manuel Sánchez-de-la-Torre, Cristina Esquinas, Camino Pérez-Gutierrez, Bartolomé Burguera, Ferran Barbé, Antonia Barceló. 2011. NADPH oxidase p22phox polymorphisms and oxidative stress in patients with obstructive sleep apnoea. *Respiratory Medicine* . [[CrossRef](#)]
6. Domenico Del Principe , Luciana Avigliano , Isabella Savini , Maria Valeria Catani . 2011. Trans-Plasma Membrane Electron Transport in Mammals: Functional Significance in Health and Disease. *Antioxidants & Redox Signaling* **14**:11, 2289-2318. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
7. Alexander Sirker, Min Zhang, Ajay M. Shah. 2011. NADPH oxidases in cardiovascular disease: insights from in vivo models and clinical studies. *Basic Research in Cardiology* . [[CrossRef](#)]
8. Rhian M Touyz, Ana M Briones. 2011. Reactive oxygen species and vascular biology: implications in human hypertension. *Hypertension Research* **34**:1, 5-14. [[CrossRef](#)]
9. Roberto Schreiber, Maria C Ferreira-Sae, Juliana A Ronchi, Jose A Pio-Magalhaes, Jose A Cipolli, Jose R Matos-Souza, Jose G Mill, Anibal E Vercesi, Jose E Krieger, Kleber G Franchini, Alexandre C Pereira, Wilson Nadruz Junior. 2011. The C242T polymorphism of the p22-phox gene (CYBA) is associated with higher left ventricular mass in Brazilian hypertensive patients. *BMC Medical Genetics* **12**:1, 114. [[CrossRef](#)]
10. Nina Queisser, Gholamreza Fazeli, Nicole Schupp. 2010. Superoxide anion and hydrogen peroxide-induced signaling and damage in angiotensin II and aldosterone action. *Biological Chemistry* **391**:11, 1265-1279. [[CrossRef](#)]
11. Anderson Saranz Zago, Leonardo Reis Silveira, Eduardo Kokubun. 2010. Effects of aerobic exercise on the blood pressure, oxidative stress and eNOS gene polymorphism in pre-hypertensive older people. *European Journal of Applied Physiology* **110**:4, 825-832. [[CrossRef](#)]
12. Panagiotis Xaplanteris, Charalambos Vlachopoulos, Katerina Baou, Carmen Vassiliadou, Ioanna Dima, Nikolaos Ioakeimidis, Christodoulos Stefanadis. 2010. The effect of p22phox -930A/G, A640G and C242T polymorphisms of NADPH oxidase on peripheral and central pressures in healthy, normotensive individuals. *Hypertension Research* **33**:8, 814-818. [[CrossRef](#)]
13. Zahra Fatehi-Hassanabad, Catherine B. Chan, Brian L. Furman. 2010. Reactive oxygen species and endothelial function in diabetes. *European Journal of Pharmacology* **636**:1-3, 8-17. [[CrossRef](#)]
14. Tomasz J. Guzik , Kathy K. Griendling . 2009. NADPH Oxidases: Molecular Understanding Finally Reaching the Clinical Level?. *Antioxidants & Redox Signaling* **11**:10, 2365-2370. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
15. Emmanuel S. Androulakis, Dimitris Tousoulis, Nikolaos Papageorgiou, Costas Tsioufis, Ioannis Kallikazaros, Christodoulos Stefanadis. 2009. Essential Hypertension. *Cardiology in Review* **17**:5, 216-221. [[CrossRef](#)]
16. Lorenzo Loffredo, Francesco Violi. 2009. The Role of Nicotinamide Adenine Dinucleotide Phosphate Oxidase in the Pathogenesis of Hypertension. *High Blood Pressure & Cardiovascular Prevention* **16**:3, 87-92. [[CrossRef](#)]
17. DEBORAH L. FEAIRHELLER, MICHAEL D. BROWN, JOON-YOUNG PARK, TINA E. BRINKLEY, SAMAR BASU, JAMES M. HAGBERG, ROBERT E. FERRELL, NICOLA M. FENTY-STEWART. 2009. Exercise Training, NADPH Oxidase p22phox Gene Polymorphisms, and Hypertension. *Medicine & Science in Sports & Exercise* **41**:7, 1421-1428. [[CrossRef](#)]
18. Hossein A. Ghofrani, Robyn J. Barst, Raymond L. Benza, Hunter C. Champion, Karen A. Fagan, Friedrich Grimminger, Marc Humbert, Gérald Simonneau, Duncan J. Stewart, Carlo Ventura, Lewis J. Rubin. 2009. Future Perspectives for the Treatment of Pulmonary Arterial Hypertension. *Journal of the American College of Cardiology* **54**:1, S108-S117. [[CrossRef](#)]

19. Moo Yeol Lee , Kathy K. Griendling . 2008. Redox Signaling, Vascular Function, and Hypertension. *Antioxidants & Redox Signaling* **10**:6, 1045-1059. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
20. Mar??a U Moreno, Gorka San Jos??, Ana Fortu??o, Oscar Beloqui, Josep Red??n, Felipe J Chaves, Dolores Corella, Javier D??ez, Guillermo Zalba. 2007. A novel CYBA variant, the ???675A/T polymorphism, is associated with essential hypertension. *Journal of Hypertension* **25**:8, 1620-1626. [[CrossRef](#)]
21. Karl-Heinz Krause. 2007. Aging: A revisited theory based on free radicals generated by NOX family NADPH oxidases. *Experimental Gerontology* **42**:4, 256-262. [[CrossRef](#)]
22. Qing Lan, Tongzhang Zheng, Min Shen, Yawei Zhang, Sophia S. Wang, Shelia H. Zahm, Theodore R. Holford, Brian Leaderer, Peter Boyle, Stephen Chanock. 2007. Genetic polymorphisms in the oxidative stress pathway and susceptibility to non-Hodgkin lymphoma. *Human Genetics* **121**:2, 161-168. [[CrossRef](#)]
23. Guillermo Zalba, Ana Fortu&ntilde;o, Gorka San Jos&eacute;, Mar&iacute;a U. Moreno, Oscar Beloqui, Javier D&iacute;ez. 2007. Oxidative Stress, Endothelial Dysfunction and Cerebrovascular Disease. *Cerebrovascular Diseases* **24**:1, 24-29. [[CrossRef](#)]
24. A. Manea, S. A. Manea, A. V. Gafencu, M. Raicu. 2007. Regulation of NADPH oxidase subunit p22 phox by NF-kB in human aortic smooth muscle cells. *Archives Of Physiology And Biochemistry* **113**:4-5, 163-172. [[CrossRef](#)]
25. Pritmohinder S. Gill , Christopher S. Wilcox . 2006. NADPH Oxidases in the Kidney. *Antioxidants & Redox Signaling* **8**:9-10, 1597-1607. [[Abstract](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
26. Shaoping Xu, Rhian M. Touyz. 2006. Reactive oxygen species and vascular remodelling in hypertension: Still alive. *Canadian Journal of Cardiology* **22**:11, 947-951. [[CrossRef](#)]
27. Natalie C Ward, Kevin D Croft. 2006. HYPERTENSION AND OXIDATIVE STRESS. *Clinical and Experimental Pharmacology and Physiology* **33**:9, 872-876. [[CrossRef](#)]
28. María U Moreno, Gorka San José, Ana Fortuño, Óscar Beloqui, Javier Díez, Guillermo Zalba. 2006. The C242T CYBA polymorphism of NADPH oxidase is associated with essential hypertension. *Journal of Hypertension* **24**:7, 1299-1306. [[CrossRef](#)]
29. Marta Ruiz-Ortega, Vanesa Esteban, Mónica Rupérez, Elsa Sánchez-López, Juan Rodríguez-Vita, Gisselle Carvajal, Jesús Egido. 2006. Renal and vascular hypertension-induced inflammation: role of angiotensin II. *Current Opinion in Nephrology and Hypertension* **15**:2, 159-166. [[CrossRef](#)]
30. Marta Ruiz-Ortega , Alberto Ortiz . 2005. Angiotensin II and Reactive Oxygen Species. *Antioxidants & Redox Signaling* **7**:9-10, 1258-1260. [[Citation](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]